

From: McLaughlin, Ryan
Sent: Tuesday, August 20, 2019 10:25 AM
To: McCleary, Jessie May
Cc: [REDACTED] 38; or.ocv.alert
Subject: Re: IACUC Training Instructions for AST-O

Thanks Jessie.

The IACUC protocols [REDACTED] 38 will work under are 6208 and 6279.

Ryan J. McLaughlin, PhD
Assistant Professor
Department of Integrative Physiology & Neuroscience
College of Veterinary Medicine
Washington State University
Pullman, WA 99164-7620

Phone: (509) 335-6448

On Aug 20, 2019, at 10:20 AM, McCleary, Jessie May <jmccleary@wsu.edu> wrote:

Hello [REDACTED] 38

I am a LVT from the Office of the Campus Veterinarian. Please let me know when you have completed the completed the AST-O training and we can get you scheduled for the hands-on portion of the training (AST-H).

Here is the link to the days available in Aug: <https://iacuc.wsu.edu/hands-on-training/>

Could you please send me the ASAF number for the protocol you are working under?

I have attached the outline of the training for you to look over and a map to our building. Let me know if you have any questions. I am looking forward to meeting with you.

Thank you,

Jessie McCleary, LVT

Washington State University

Office of the Campus Veterinarian

jmccleary@wsu.edu

Office (509)335-2595

Main line (509)335-6246

Emergency phone (509)330-1871

From: IACUC <or.ora.iacuc@wsu.edu>

Sent: Tuesday, August 20, 2019 10:09 AM

To: [REDACTED] 38

Cc: McLaughlin, Ryan <ryan.mclaughlin@wsu.edu>; or.ocv.alert <or.ocv.alert@wsu.edu>

Subject: IACUC Training Instructions for AST-O

Importance: High

Hi [REDACTED] 38

This is a courtesy notice that you are required for Dr. McLaughlin's ASAFs to complete the ***Aseptic Surgical Technique WSU (AST-O)***, an online course on AALAS Learning Library website that is required by WSU IACUC for all new researchers/ faculty/ staff performing surgery on research or teaching animals. Board-certified surgeons working within their specialty may be exempt.

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Our records indicate that you have an account with AALAS Learning Library.

- The link to log in is: <https://aalaslearninglibrary.org/index.html#/login/signin>
- Enter your User name: 41
- Enter your password. If you need to reset your password, you can click on the “I forgot my password” link. If that does not work, contact the Animal Welfare Program.

Step 1: Once you log on, you will be directed to the AALAS homepage. Complete your profile located in the upper right corner with the person icon (arrow 1 in Figure 1).

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Once you have passed the exam, you have completed the course.

Figure 1:

<image001.png>

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(DVM, LVT, MD, and RN may be exempt from the hands-on course.)

If you have any issues regarding IACUC training, please contact the Animal Welfare Program at (509) 335-5353 or email at iacuc@wsu.edu.

Sincerely,

Ashley Williams

Animal Welfare Program

Washington State University | Office of Research Assurances

Neill Hall 431 | Pullman, WA 99164-3143

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<https://iacuc.wsu.edu>

<AST-H Rodents Outline 2018.docx><OCV Location.pptx>

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<https://iacuc.wsu.edu>

Aseptic Surgical Technique for Rodents

Hands-on Workshop

Washington State University
Office of the Campus Veterinarian-Vet Services

1. Introduction

- a. Discuss similarities & differences between rodent and non-rodent surgery
 - i. Similar- Both require asepsis for anything touching the surgical area
 - ii. Differences are primarily due to the small size of the rodent
 - 1. Designated space vs. dedicated facility (room or suite)
 - a. Clean lab bench in non-traffic area
 - 2. Clean lab coat vs. sterile surgical gown
 - 3. Work alone vs. surgical team
 - a. Without assistant, asepsis requires careful planning

2. Procedure Area Preparation

- a. Designated for surgery currently, no traffic/ drafts, post signs “Aseptic Surgery in Progress”
- b. Surgical/anesthesia records and disinfected pen
- c. Disinfect surfaces. Remove unneeded items.
- d. Animal Prep area: clippers, scrub solution, eye ointment, disposable layer
- e. Surgical Area: heat source, light, glass bead sterilizer? Absorbent clean or sterile layer, sterile instrument pack, drape and suture, (wait to open until just before gloves on)
- f. Post-operative area: heat source on lowest setting, under half of recovery cage, white pad or paper towels (instead of bedding), analgesic and saline drawn into syringes(warm syringes with heating pad)

3. Instruments and Materials Preparation

- a. Autoclaving- steam heat (moist heat, physical method)
 - i. Metal instruments, some catheter tubing, saline, water
 - ii. Packaging
 - 1. Surgical packs- gauze, drape, instruments
 - 2. Envelopes- Visi-peel
 - 3. Indicator tape, indicator strips
 - 4. Monitor autoclave quarterly with biological indicator
 - iii. Date autoclaved materials – maximum 6 month expiration
 - iv. Pre-packaged sterile supplies- suture, blades (check expiration dates)
- b. Glass bead sterilizers- dry heat (modified aseptic technique “tips only”)
 - i. Start with autoclaved pack then use hot glass beads between animals
 - ii. Wipe instruments to remove blood & debris before use

Revised 9/21/18

Obtained by Rise for Animals.
Uploaded to Animal Research Laboratory Overview (ARLO) on 02/03/2021

- iii. Instrument tips in heat for 30-60 seconds (too long may warp)
 - iv. Cool before use to prevent burns to animal or surgeon
 - v. New autoclave pack every 5 animals.
 - vi. Only tips are re-sterilized. Position: tips away/ handles toward surgeon
 - vii. New surgery gloves for each animal (unless assistant prepped new animal)
 - viii. Tips only surgical technique (handles & hands are clean but not sterile)
- c. Chemical sterilants- liquid “cold sterilization”
- i. Liquid sterilants require 6-8 hours contact times to kill spores
 - ii. Liquid must contact all surfaces (fill tubing)
 - iii. Rinse chemicals off with sterile saline/H₂O before use
 - iv. Alcohol is a disinfectant but not a sterilant
- d. Chemical sterilants- gas “cold and dry sterilization”
- i. Ethylene Oxide
 - ii. Vaporized Hydrogen Peroxide

4. Animal Preparation

- a. Anesthesia
- i. Injectable or inhalation- assemble needed supplies (DEA regulations)
 - ii. Weigh each animal pre-operatively & write in surgery/anesthesia record (grams)
 - iii. Calculate dosages and draw up anesthesia, analgesia,
 - iv. Give subcutaneous saline before the surgery if possible
 - v. No food or water restriction for rodents
 - vi. Rodents are susceptible to dehydration, hypoglycemia, hypothermia
- b. Eye ointment
- i. Anesthetized rodents don't close eyes
 - ii. Prevent drying of corneas
 - iii. Use ophthalmic ointment (artificial tears)
- c. Monitor anesthetic depth (variation in strains and individual animals)
- i. Observe respiratory rate and depth (Count 15 seconds X 4)
 - ii. Color of mucus membranes, ears, tail, feet
 - iii. Toe pinch withdrawal reflex? Use fingers or hemostat
 - 1. If withdraw, too light for surgery but clipping is OK
 - 2. Intra-operative- Maintain sterility using hemostats- handles sterile
 - iv. If using injectable anesthetic (ketamine/xylazine) only boost with ketamine (1/3rd).
- d. Clip hair
- i. Blades #40 - maintain with spray cleaner/lubricant
 - ii. Razor or Depilatory Cream (Nair) may be used

- e. Scrub: can be done with exam gloves
 - i. Chlorhexidine (scrub or solution) then alcohol OR Povidone/iodine (scrub or solution) then alcohol
 - ii. Start at center and move out in spiral to clipped margins
 - iii. Triple scrub then may apply solution to dry on (not scrub- contains soap)
 - iv. Cotton swabs versus gauze sponges
 - v. Don't soak animal or get disinfectant in eyes

5. Surgeon Preparation

- a. Required
 - i. Remove earrings, necklaces, rings, watches, scarves, etc...
 - ii. Clean lab coat
 - 1. Free of debris, hair, or bedding to fall into surgical area
 - 2. Snug lab coat cuffs or tape around wide sleeves or roll up sleeves
 - iii. Face mask
 - 1. protect surgical field from bacteria (surgeon's exhaled breath)
 - 2. Mask is clean but not sterile (Do not touch with gloved hands)
 - iv. Surgical gloves – Wait until animal is prepped & ready to be draped
Working alone? Before gloving ask yourself: **LIST** (Light on, Instrument pack open, Supplies/suture, Toe pinch check anesthesia/ Time starting noted)
 - 1. Individually packaged latex or nitrile surgical gloves
 - a. **NOT** exam gloves even if cleaned and/or autoclaved
 - b. Do not re-sterilize & re-use surgical gloves
 - c. Surgical gloves- check expiration date
 - d. Sized in half sizes to fit (example: 6.0, 6.5, 7.0, 7.5)
 - 2. Wash hands with soap and warm water. Dry with clean towel.
 - 3. Aseptic gloving technique- demonstrate & practice
 - 4. If gloves are contaminated during surgery– get new gloves
 - 5. Put on new sterile gloves every 3 animals or if they become contaminated between surgeries.
 - v. **Review your ASAF** surgery section on “How aseptic technique is maintained”
You must adhere to what is written in the ASAF.
- b. Recommended
 - i. Hair cap/ bonnet recommended. Tie back long hair. Beard cover.
 - ii. An Assistant: the surgeon can't touch anything non-sterile once gloved. Surgery is more efficient, faster and less complicated if there is an assistant.
 - iii. **Review ASAF**, if cap for example, is listed then need to wear.

6. Draping

- a. Open sterile instrument and drape pack → put on sterile gloves → then may handle drape

- b. Place drape & lift drape to check toe pinch with hemostats without contaminating sterile gloves.
- c. Alternative: Press N' Seal plastic wrap –use will be demonstrated at workshop

7. Perform surgery

- a. Intraoperative monitoring of anesthetic depth- surgical record
- b. Maintain sterile gloves and sterile field
- c. Batch surgery? Working alone? Heat sterilize instrument tips prior to handling animal.

8. Recovery & post-operative care

- a. Post-operative analgesic and warm saline
- b. Wrap rodent in huck or paper towel to maintain body heat
- c. Rotate position every 10 minutes until sternal, semi awake
- d. Maintain heat source & keep separate until animal is moving about cage
- e. Check surgery rodents end of first day
- f. Monitor twice daily for first 3 days, then once daily for appetite, activity, grooming
- g. Check incision for redness, swelling, discharge, suture failure
- h. Water-soaked kibble or Gel pack in cage bottom advised
- i. Document observations, analgesics, and date of suture removal (~10 days)
- j. Analgesia discussion: do what is in approved protocol

9. Conclusion

- a. Success requires careful planning & preparation
- b. Difficult to do surgery alone especially on multiple animals- recommend having assistant to help prepare and recover animals while surgeon focuses on surgery

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From: McCleary, Jessie May
Sent: Tuesday, August 20, 2019 8:23 AM
To: 38
Cc: Fuchs, Rita; or.ocv.alert
Subject: RE: IACUC Training Instructions for AST-O
Attachments: AST-H Rodents Outline 2018.docx; OCV Location.pptx

Hello 38

Thank you for completing the on-line training.

I have an Aseptic Surgical Technique Hands-on workshop scheduled for Tues the 27th 10am-12pm. Please let me know if this works for you.

I have attached the outline of the training for you to look over and a map to our building. Let me know if you have any questions. I am looking forward to meeting with you.

Thank you,

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jmccleary@wsu.edu
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From: IACUC <or.ora.iacuc@wsu.edu>
Sent: Monday, August 19, 2019 9:15 AM
To: 38
Cc: Fuchs, Rita <rita.fuchs@wsu.edu>; or.ocv.alert <or.ocv.alert@wsu.edu>
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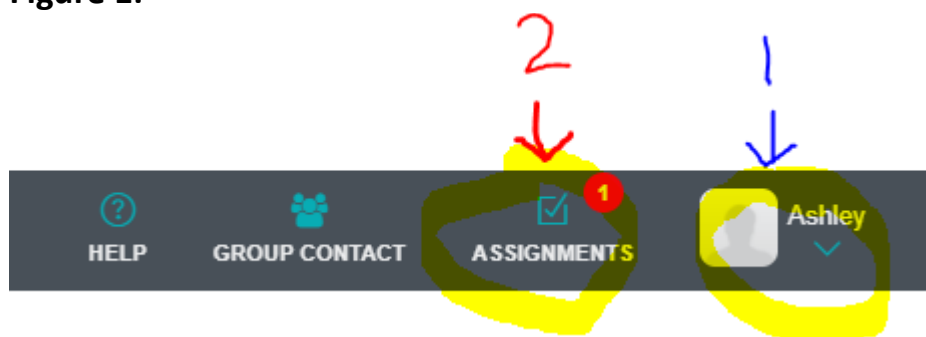
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Sent: Thursday, August 15, 2019 5:02 PM
To: Rossi, David; [REDACTED] 38
Cc: or.ocv.alert
Subject: RE: IACUC Training Instructions for AST-O

Thank you Dr Rossi, I will look into what we need to do with proceeding with [REDACTED] 38 training.
Jess

From: Rossi, David <david.rossi@wsu.edu>
Sent: Thursday, August 15, 2019 4:49 PM
To: McCleary, Jessie May <jmccleary@wsu.edu>; [REDACTED] 38
Cc: or.ocv.alert <or.ocv.alert@wsu.edu>
Subject: RE: IACUC Training Instructions for AST-O

Dear Jessie and [REDACTED] 38

[REDACTED] 38 is actually starting a rotation in the laboratory of Dr. Jim Peters, so will likely not be doing anything in my laboratory for about 8 weeks. Obviously at that time or in advance she'll need to be added to my protocol, so here are the two relevant protocols:

Breeding 6285

Usage 4979

But you should work with [REDACTED] 38 to get her onto Jim Peters IACUC protocols for the nearer term situation, unless I am confused about something?

Let me know if you need anything else from me now.

Thanks,

David

PS. [REDACTED] 38 as you do any training etc. that needs to be associated with IACUC numbers, please include my two, above, along with whatever you need to do for Jim Peters.

David Rossi PhD
Associate Professor
Department of Integrative Physiology and Neuroscience
1815 Ferdinand's Lane
Washington State University
Pullman, WA 99164-7620
Phone: 509-335-7671
e-mail: David.Rossi@wsu.edu

From: McCleary, Jessie May
Sent: Thursday, August 15, 2019 4:41 PM
To: [REDACTED] 38
Cc: Rossi, David <david.rossi@wsu.edu>; or.ocv.alert <or.ocv.alert@wsu.edu>
Subject: RE: IACUC Training Instructions for AST-O

Hello [REDACTED] 38

I am a LVT from the Office of the Campus Veterinarian. Please let me know when you have completed the completed the AST-O training and we can get you scheduled for the hands-on portion of the training (AST-H).

Here is the link to the days available in August: <https://iacuc.wsu.edu/hands-on-training/>

Could you please send me the ASAF number for the protocol you are working under?

I have attached the outline of the training for you to look over and a map to our building. Let me know if you have any questions. I am looking forward to meeting with you.

Thank you,

Jessie McCleary, LVT
Washington State University
Office of the Campus Veterinarian
jmccleary@wsu.edu
Office (509)335-2595
Main line (509)335-6246
Emergency phone (509)330-1871

From: IACUC <or.ora.iacuc@wsu.edu>

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Importance: High

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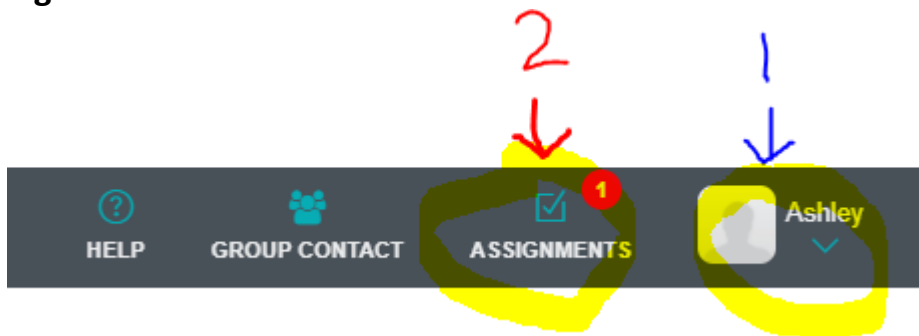
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Sincerely,

Ashley Williams

Animal Welfare Program
Washington State University | Office of Research Assurances
Neill Hall 431 | Pullman, WA 99164-3143
iacuc@wsu.edu | Phone: (509) 335-5353
<https://iacuc.wsu.edu>

From: Rossi, David
Sent: Thursday, August 15, 2019 4:49 PM
To: McCleary, Jessie May; [REDACTED] 38
Cc: or.ocv.alert
Subject: RE: IACUC Training Instructions for AST-O

Dear Jessie and [REDACTED] 38

[REDACTED] 38 is actually starting a rotation in the laboratory of Dr. Jim Peters, so will likely not be doing anything in my laboratory for about 8 weeks. Obviously at that time or in advance she'll need to be added to my protocol, so here are the two relevant protocols:

Breeding 6285
Usage 4979

But you should work with [REDACTED] 38 to get her onto Jim Peters IACUC protocols for the nearer term situation, unless I am confused about something?

Let me know if you need anything else from me now.

Thanks,
David

PS. [REDACTED] 38 as you do any training etc. that needs to be associated with IACUC numbers, please include my two, above, along with whatever you need to do for Jim Peters.

David Rossi PhD
Associate Professor
Department of Integrative Physiology and Neuroscience
1815 Ferdinand's Lane
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Aseptic Surgical Technique for Rodents

Hands-on Workshop

Washington State University
Office of the Campus Veterinarian-Vet Services

1. Introduction

- a. Discuss similarities & differences between rodent and non-rodent surgery
 - i. Similar- Both require asepsis for anything touching the surgical area
 - ii. Differences are primarily due to the small size of the rodent
 - 1. Designated space vs. dedicated facility (room or suite)
 - a. Clean lab bench in non-traffic area
 - 2. Clean lab coat vs. sterile surgical gown
 - 3. Work alone vs. surgical team
 - a. Without assistant, asepsis requires careful planning

2. Procedure Area Preparation

- a. Designated for surgery currently, no traffic/ drafts, post signs “Aseptic Surgery in Progress”
- b. Surgical/anesthesia records and disinfected pen
- c. Disinfect surfaces. Remove unneeded items.
- d. Animal Prep area: clippers, scrub solution, eye ointment, disposable layer
- e. Surgical Area: heat source, light, glass bead sterilizer? Absorbent clean or sterile layer, sterile instrument pack, drape and suture, (wait to open until just before gloves on)
- f. Post-operative area: heat source on lowest setting, under half of recovery cage, white pad or paper towels (instead of bedding), analgesic and saline drawn into syringes(warm syringes with heating pad)

3. Instruments and Materials Preparation

- a. Autoclaving- steam heat (moist heat, physical method)
 - i. Metal instruments, some catheter tubing, saline, water
 - ii. Packaging
 - 1. Surgical packs- gauze, drape, instruments
 - 2. Envelopes- Visi-peel
 - 3. Indicator tape, indicator strips
 - 4. Monitor autoclave quarterly with biological indicator
 - iii. Date autoclaved materials – maximum 6 month expiration
 - iv. Pre-packaged sterile supplies- suture, blades (check expiration dates)
- b. Glass bead sterilizers- dry heat (modified aseptic technique “tips only”)
 - i. Start with autoclaved pack then use hot glass beads between animals
 - ii. Wipe instruments to remove blood & debris before use

Revised 9/21/18

Obtained by Rise for Animals.
Uploaded to Animal Research Laboratory Overview (ARLO) on 02/03/2021

- iii. Instrument tips in heat for 30-60 seconds (too long may warp)
 - iv. Cool before use to prevent burns to animal or surgeon
 - v. New autoclave pack every 5 animals.
 - vi. Only tips are re-sterilized. Position: tips away/ handles toward surgeon
 - vii. New surgery gloves for each animal (unless assistant prepped new animal)
 - viii. Tips only surgical technique (handles & hands are clean but not sterile)
- c. Chemical sterilants- liquid “cold sterilization”
- i. Liquid sterilants require 6-8 hours contact times to kill spores
 - ii. Liquid must contact all surfaces (fill tubing)
 - iii. Rinse chemicals off with sterile saline/H₂O before use
 - iv. Alcohol is a disinfectant but not a sterilant
- d. Chemical sterilants- gas “cold and dry sterilization”
- i. Ethylene Oxide
 - ii. Vaporized Hydrogen Peroxide

4. Animal Preparation

- a. Anesthesia
- i. Injectable or inhalation- assemble needed supplies (DEA regulations)
 - ii. Weigh each animal pre-operatively & write in surgery/anesthesia record (grams)
 - iii. Calculate dosages and draw up anesthesia, analgesia,
 - iv. Give subcutaneous saline before the surgery if possible
 - v. No food or water restriction for rodents
 - vi. Rodents are susceptible to dehydration, hypoglycemia, hypothermia
- b. Eye ointment
- i. Anesthetized rodents don't close eyes
 - ii. Prevent drying of corneas
 - iii. Use ophthalmic ointment (artificial tears)
- c. Monitor anesthetic depth (variation in strains and individual animals)
- i. Observe respiratory rate and depth (Count 15 seconds X 4)
 - ii. Color of mucus membranes, ears, tail, feet
 - iii. Toe pinch withdrawal reflex? Use fingers or hemostat
 - 1. If withdraw, too light for surgery but clipping is OK
 - 2. Intra-operative- Maintain sterility using hemostats- handles sterile
 - iv. If using injectable anesthetic (ketamine/xylazine) only boost with ketamine (1/3rd).
- d. Clip hair
- i. Blades #40 - maintain with spray cleaner/lubricant
 - ii. Razor or Depilatory Cream (Nair) may be used

- e. Scrub: can be done with exam gloves
 - i. Chlorhexidine (scrub or solution) then alcohol OR Povidone/iodine (scrub or solution) then alcohol
 - ii. Start at center and move out in spiral to clipped margins
 - iii. Triple scrub then may apply solution to dry on (not scrub- contains soap)
 - iv. Cotton swabs versus gauze sponges
 - v. Don't soak animal or get disinfectant in eyes

5. Surgeon Preparation

- a. Required
 - i. Remove earrings, necklaces, rings, watches, scarves, etc...
 - ii. Clean lab coat
 - 1. Free of debris, hair, or bedding to fall into surgical area
 - 2. Snug lab coat cuffs or tape around wide sleeves or roll up sleeves
 - iii. Face mask
 - 1. protect surgical field from bacteria (surgeon's exhaled breath)
 - 2. Mask is clean but not sterile (Do not touch with gloved hands)
 - iv. Surgical gloves – Wait until animal is prepped & ready to be draped
Working alone? Before gloving ask yourself: **LIST** (Light on, Instrument pack open, Supplies/suture, Toe pinch check anesthesia/ Time starting noted)
 - 1. Individually packaged latex or nitrile surgical gloves
 - a. **NOT** exam gloves even if cleaned and/or autoclaved
 - b. Do not re-sterilize & re-use surgical gloves
 - c. Surgical gloves- check expiration date
 - d. Sized in half sizes to fit (example: 6.0, 6.5, 7.0, 7.5)
 - 2. Wash hands with soap and warm water. Dry with clean towel.
 - 3. Aseptic gloving technique- demonstrate & practice
 - 4. If gloves are contaminated during surgery– get new gloves
 - 5. Put on new sterile gloves every 3 animals or if they become contaminated between surgeries.
 - v. **Review your ASAF** surgery section on “How aseptic technique is maintained”
You must adhere to what is written in the ASAF.
- b. Recommended
 - i. Hair cap/ bonnet recommended. Tie back long hair. Beard cover.
 - ii. An Assistant: the surgeon can't touch anything non-sterile once gloved. Surgery is more efficient, faster and less complicated if there is an assistant.
 - iii. **Review ASAF**, if cap for example, is listed then need to wear.

6. Draping

- a. Open sterile instrument and drape pack → put on sterile gloves → then may handle drape

- b. Place drape & lift drape to check toe pinch with hemostats without contaminating sterile gloves.
- c. Alternative: Press N' Seal plastic wrap –use will be demonstrated at workshop

7. Perform surgery

- a. Intraoperative monitoring of anesthetic depth- surgical record
- b. Maintain sterile gloves and sterile field
- c. Batch surgery? Working alone? Heat sterilize instrument tips prior to handling animal.

8. Recovery & post-operative care

- a. Post-operative analgesic and warm saline
- b. Wrap rodent in huck or paper towel to maintain body heat
- c. Rotate position every 10 minutes until sternal, semi awake
- d. Maintain heat source & keep separate until animal is moving about cage
- e. Check surgery rodents end of first day
- f. Monitor twice daily for first 3 days, then once daily for appetite, activity, grooming
- g. Check incision for redness, swelling, discharge, suture failure
- h. Water-soaked kibble or Gel pack in cage bottom advised
- i. Document observations, analgesics, and date of suture removal (~10 days)
- j. Analgesia discussion: do what is in approved protocol

9. Conclusion

- a. Success requires careful planning & preparation
- b. Difficult to do surgery alone especially on multiple animals- recommend having assistant to help prepare and recover animals while surgeon focuses on surgery

OCV is located in the LARC Building (in the yellow circle), the orange star is where the entrance door to the building. Front office is on your left. Josh our receptionist will bring you to Training Room. If he is not at his desk ring the bell on the wall for assistance. (The blue star is the training room #198.)



From: McCleary, Jessie May
Sent: Tuesday, August 27, 2019 9:23 AM
To: [REDACTED] 38
Cc: Quock, Raymond; or.ocv.alert
Subject: RE: IACUC Training Instructions for AST-O
Attachments: AST-H Rodents Outline 2018.docx; OCV Location.pptx

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- Enter your User name: 41
- Enter your password. If you need to reset your password, you can click on the “I forgot my password” link. If that does not work, contact the Animal Welfare Program.

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- d. Animal Prep area: clippers, scrub solution, eye ointment, disposable layer
- e. Surgical Area: heat source, light, glass bead sterilizer? Absorbent clean or sterile layer, sterile instrument pack, drape and suture, (wait to open until just before gloves on)
- f. Post-operative area: heat source on lowest setting, under half of recovery cage, white pad or paper towels (instead of bedding), analgesic and saline drawn into syringes(warm syringes with heating pad)

3. Instruments and Materials Preparation

- a. Autoclaving- steam heat (moist heat, physical method)
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 - ii. Packaging
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 - iv. If using injectable anesthetic (ketamine/xylazine) only boost with ketamine (1/3rd).
- d. Clip hair
- i. Blades #40 - maintain with spray cleaner/lubricant
 - ii. Razor or Depilatory Cream (Nair) may be used

- e. Scrub: can be done with exam gloves
 - i. Chlorhexidine (scrub or solution) then alcohol OR Povidone/iodine (scrub or solution) then alcohol
 - ii. Start at center and move out in spiral to clipped margins
 - iii. Triple scrub then may apply solution to dry on (not scrub- contains soap)
 - iv. Cotton swabs versus gauze sponges
 - v. Don't soak animal or get disinfectant in eyes

5. Surgeon Preparation

- a. Required
 - i. Remove earrings, necklaces, rings, watches, scarves, etc...
 - ii. Clean lab coat
 - 1. Free of debris, hair, or bedding to fall into surgical area
 - 2. Snug lab coat cuffs or tape around wide sleeves or roll up sleeves
 - iii. Face mask
 - 1. protect surgical field from bacteria (surgeon's exhaled breath)
 - 2. Mask is clean but not sterile (Do not touch with gloved hands)
 - iv. Surgical gloves – Wait until animal is prepped & ready to be draped
Working alone? Before gloving ask yourself: **LIST** (Light on, Instrument pack open, Supplies/suture, Toe pinch check anesthesia/ Time starting noted)
 - 1. Individually packaged latex or nitrile surgical gloves
 - a. **NOT** exam gloves even if cleaned and/or autoclaved
 - b. Do not re-sterilize & re-use surgical gloves
 - c. Surgical gloves- check expiration date
 - d. Sized in half sizes to fit (example: 6.0, 6.5, 7.0, 7.5)
 - 2. Wash hands with soap and warm water. Dry with clean towel.
 - 3. Aseptic gloving technique- demonstrate & practice
 - 4. If gloves are contaminated during surgery– get new gloves
 - 5. Put on new sterile gloves every 3 animals or if they become contaminated between surgeries.
 - v. **Review your ASAF** surgery section on “How aseptic technique is maintained”
You must adhere to what is written in the ASAF.
- b. Recommended
 - i. Hair cap/ bonnet recommended. Tie back long hair. Beard cover.
 - ii. An Assistant: the surgeon can't touch anything non-sterile once gloved. Surgery is more efficient, faster and less complicated if there is an assistant.
 - iii. **Review ASAF**, if cap for example, is listed then need to wear.

6. Draping

- a. Open sterile instrument and drape pack → put on sterile gloves → then may handle drape

- b. Place drape & lift drape to check toe pinch with hemostats without contaminating sterile gloves.
- c. Alternative: Press N' Seal plastic wrap –use will be demonstrated at workshop

7. Perform surgery

- a. Intraoperative monitoring of anesthetic depth- surgical record
- b. Maintain sterile gloves and sterile field
- c. Batch surgery? Working alone? Heat sterilize instrument tips prior to handling animal.

8. Recovery & post-operative care

- a. Post-operative analgesic and warm saline
- b. Wrap rodent in huck or paper towel to maintain body heat
- c. Rotate position every 10 minutes until sternal, semi awake
- d. Maintain heat source & keep separate until animal is moving about cage
- e. Check surgery rodents end of first day
- f. Monitor twice daily for first 3 days, then once daily for appetite, activity, grooming
- g. Check incision for redness, swelling, discharge, suture failure
- h. Water-soaked kibble or Gel pack in cage bottom advised
- i. Document observations, analgesics, and date of suture removal (~10 days)
- j. Analgesia discussion: do what is in approved protocol

9. Conclusion

- a. Success requires careful planning & preparation
- b. Difficult to do surgery alone especially on multiple animals- recommend having assistant to help prepare and recover animals while surgeon focuses on surgery

OCV is located in the LARC Building (in the yellow circle), the orange star is where the entrance door to the building. Front office is on your left. Josh our receptionist will bring you to Training Room. If he is not at his desk ring the bell on the wall for assistance. (The blue star is the training room #198.)



From: Luft, Jan Elizabeth
Sent: Monday, May 20, 2019 4:28 PM
To: Gorence, Galen Jay; Anderson, Gwen Marie; or.ocv.alert; Gabelmann, Doris L
Subject: RE: LARC Power outage tomorrow AM

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I do have doctor's appt at 8 but it should not take too long.

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From: Gold, Jenifer Robin
Sent: Friday, May 10, 2019 8:13 AM
To: Clyde, Gaylynn Goolsby; Turner, Gary Dean; Woodford, Nina Lynne
Cc: or.ocv.alert
Subject: Re: Message from "RNP002673B143C5"

Gaylynn,

33 said she could get Shimmer in a week. So let's plan on examination next week. I believe she is staying in WA but will double check!

Jen

Jenifer R. Gold DVM Diplomate ACVIM, ACVECC
Clinical Associate Professor
Department of Veterinary Clinical Sciences
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Pullman, WA. 99164
(509) 432-6764

Love recognizes no barriers. It jumps, hurdles, leaps fences, penetrates walls to arrive at its destination full of hope-Maya Angelou

From: Clyde, Gaylynn Goolsby
Sent: Thursday, May 9, 2019 8:06:25 AM
To: Turner, Gary Dean; Gold, Jenifer Robin; Woodford, Nina Lynne
Cc: or.ocv.alert
Subject: RE: Message from "RNP002673B143C5"

Hi everyone,

I can perform physical exam for adoption. If she is crossing state lines more than likely she will need Coggins test, CVI and brand inspection which I could do also at the time of the PE if you would like?

Let me know time frame for pick up and I will get her on the schedule.

Thank you,

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Phone (509) 335-4991 or (509) 335-6246
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Email: gclyde@wsu.edu
Program URL: <http://campusvet.wsu.edu>

From: Turner, Gary Dean <gdtturner@wsu.edu>
Sent: Thursday, May 9, 2019 7:35 AM
To: Gold, Jenifer Robin <jgold@wsu.edu>; Woodford, Nina Lynne <nwoodford@wsu.edu>
Cc: Clyde, Gaylynn Goolsby <gclyde@wsu.edu>
Subject: RE: Message from "RNP002673B143C5"

Also...she has blue collar #104 for reference. Her MR# is on her transfer paperwork.

Gary D. Turner BPS
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Cc: Clyde, Gaylynn Goolsby; Turner, Gary Dean
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I'd also like to pull & read her medical record for the health history

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From: Gold, Jenifer Robin <jgold@wsu.edu>
Sent: Wednesday, May 8, 2019 11:57 AM
To: Woodford, Nina Lynne <nwoodford@wsu.edu>
Subject: Re: Message from "RNP002673B143C5"

Sounds good I'll send to [REDACTED]
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Sent from my iPhone

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From: 1020scanner@vetmed.wsu.edu <1020scanner@vetmed.wsu.edu>
Sent: Wednesday, May 8, 2019 11:27 AM
To: Gold, Jenifer Robin
Subject: Message from "RNP002673B143C5"

This E-mail was sent from "RNP002673B143C5" (MP 6054).

Scan Date: 05.08.2019 11:27:31 (-0700)
Queries to: 1020scanner@vetmed.wsu.edu

From: Gold, Jenifer Robin
Sent: Thursday, May 09, 2019 8:08 AM
To: Clyde, Gaylynn Goolsby; Turner, Gary Dean; Woodford, Nina Lynne
Cc: or.ocv.alert
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Thank you! Waiting on a reply from [REDACTED] I will let you know!
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Love recognizes no barriers. It jumps, hurdles, leaps fences, penetrates walls to arrive at its destination full of hope-Maya Angelou

From: 1020scanner@vetmed.wsu.edu <1020scanner@vetmed.wsu.edu>
Sent: Wednesday, May 8, 2019 11:27 AM
To: Gold, Jenifer Robin
Subject: Message from "RNP002673B143C5"

This E-mail was sent from "RNP002673B143C5" (MP 6054).

Scan Date: 05.08.2019 11:27:31 (-0700)
Queries to: 1020scanner@vetmed.wsu.edu

From: Clyde, Gaylynn Goolsby
Sent: Thursday, May 09, 2019 8:06 AM
To: Turner, Gary Dean; Gold, Jenifer Robin; Woodford, Nina Lynne
Cc: or.ocv.alert
Subject: RE: Message from "RNP002673B143C5"

Hi everyone,

I can perform physical exam for adoption. If she is crossing state lines more than likely she will need Coggins test, CVI and brand inspection which I could do also at the time of the PE if you would like?

Let me know time frame for pick up and I will get her on the schedule.

Thank you,

Gay Lynn Clyde, DVM
Assistant Director Campus Veterinary Services
Office of Campus Veterinarian
Washington State University
Pullman, WA. 99164-1165
Phone (509) 335-4991 or (509) 335-6246
Fax (509) 335-3162
Email: gclyde@wsu.edu
Program URL: <http://campusvet.wsu.edu>

From: Turner, Gary Dean <gdturner@wsu.edu>
Sent: Thursday, May 9, 2019 7:35 AM
To: Gold, Jenifer Robin <jgold@wsu.edu>; Woodford, Nina Lynne <nwoodford@wsu.edu>
Cc: Clyde, Gaylynn Goolsby <gclyde@wsu.edu>
Subject: RE: Message from "RNP002673B143C5"

Also...she has blue collar #104 for reference. Her MR# is on her transfer paperwork.

Gary D. Turner BPS
Manager, Teaching and Research Herds Unit & VTH Animal Care
Washington State University College of Veterinary Medicine
Department of Veterinary Clinical Sciences
PO Box 646610
Pullman, WA. 99164-6610
509-335-0818 FAX 509-335-0880
gdturner@vetmed.wsu.edu
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Cc: Clyde, Gaylynn Goolsby <gclyde@wsu.edu>; Turner, Gary Dean <gdtturner@wsu.edu>
Subject: Re: Message from "RNP002673B143C5"

Sounds good..I will see if I can find a time line when 33 wants to get her.
J

Jenifer R. Gold DVM Diplomate ACVIM, ACVECC
Clinical Associate Professor
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Washington State University,
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(509) 432-6764

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From: Woodford, Nina Lynne
Sent: Wednesday, May 8, 2019 5:13:00 PM
To: Gold, Jenifer Robin
Cc: Clyde, Gaylynn Goolsby; Turner, Gary Dean
Subject: RE: Message from "RNP002673B143C5"

Sounds good. I left a message with Gary and on your cell phone to set a time for the physical exam

I'd also like to pull & read her medical record for the health history

Thanks
Nina

Nina Woodford DVM, MPH DACLAM
Director and Attending Veterinarian
Office of the Campus Veterinarian
Washington State University
509-335-6246
nwoodford@wsu.edu

From: Gold, Jenifer Robin <jgold@wsu.edu>
Sent: Wednesday, May 8, 2019 11:57 AM
To: Woodford, Nina Lynne <nwoodford@wsu.edu>
Subject: Re: Message from "RNP002673B143C5"

Sounds good I'll send to 33
Your horse sounds perfect for an anatomy horse. Yes I would like you all to do the physical exams etc!

J

Sent from my iPhone

On May 8, 2019, at 11:53 AM, Woodford, Nina Lynne <nwoodford@wsu.edu> wrote:

33 needs to fill in and sign the new owner and transport section (date can be left blank until determined). Dr. Clyde or myself or one of the VTH veterinarians not associated with the teaching herd can do the outgoing physical exam. Do you have someone in mind or would you like us to it?

I do have a 28 year old quarter-arab mare that was my son's first horse. Too small for him now and doesn't get along with the sheep, goat & donkey at home. Picks on the smaller critters.

I have meetings until 3 but can call later today

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From: Gold, Jenifer Robin <jgold@wsu.edu>
Sent: Wednesday, May 8, 2019 11:21 AM
To: Woodford, Nina Lynne <nwoodford@wsu.edu>
Subject: Fw: Message from "RNP002673B143C5"

Nina,
Shimmer is going to get adopted by **33**. I don't know if you need her to fill out the paperwork first or??
I have attached the form. Also filled out online info.
Gary told me you have a horse that we might be able to use for Anatomy lab?
Would love to talk more!
Jen

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Sent: Friday, May 10, 2019 8:17 PM
To: Clyde, Gaylynn Goolsby; Turner, Gary Dean; Woodford, Nina Lynne
Cc: or.ocv.alert
Subject: Re: Message from "RNP002673B143C5"

All,
Shimmer is going to a place in WA.
So should be easy.
J

Jenifer R. Gold DVM Diplomate ACVIM, ACVECC
Clinical Associate Professor
Department of Veterinary Clinical Sciences
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Pullman, WA. 99164
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From: Gold, Jenifer Robin
Sent: Friday, May 10, 2019 8:12:45 AM
To: Clyde, Gaylynn Goolsby; Turner, Gary Dean; Woodford, Nina Lynne
Cc: or.ocv.alert
Subject: Re: Message from "RNP002673B143C5"

Gaylynn,
[REDACTED] said she could get Shimmer in a week. So let's plan on examination next week. I believe she is staying in WA but will double check!
Jen

Jenifer R. Gold DVM Diplomate ACVIM, ACVECC
Clinical Associate Professor
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From: Clyde, Gaylynn Goolsby
Sent: Thursday, May 9, 2019 8:06:25 AM
To: Turner, Gary Dean; Gold, Jenifer Robin; Woodford, Nina Lynne
Cc: or.ocv.alert
Subject: RE: Message from "RNP002673B143C5"

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Sent from my iPhone

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Scan Date: 05.08.2019 11:27:31 (-0700)

Queries to: 1020scanner@vetmed.wsu.edu

From: Keeton, Denise
Sent: Tuesday, May 07, 2019 2:50 PM
To: Heineke, Merle J; Lele, Aaditi; Liu, David
Cc: Woodford, Nina Lynne; ocv.animal.shipping
Subject: RE: Mice marked for shipment to UMASS

Nina,
There is no further paperwork needed on our end. These mice originated from UMASS with the same PI they are going back to. No MTA to send them back.

Thanks,
Denise

Denise Keeton
Assistant Director, Business Operations
Office of Commercialization
Washington State University
Lighty 280/286
Pullman WA 99164-1060
(o) 509-335-1210 (f) 509-335-7237
ddkeeton@wsu.edu

Go Cougs!

THE INFORMATION CONTAINED IN THIS TRANSMISSION IS CONFIDENTIAL, MAY BE LEGALLY PRIVILEGED, AND IS ONLY FOR THE INTENDED RECIPIENT IDENTIFIED ABOVE. IF YOU HAVE RECEIVED THIS TRANSMISSION IN ERROR, PLEASE IMMEDIATELY NOTIFY US BY TELEPHONE, RETURN THE ORIGINAL MESSAGE TO US, AND RETAIN NO COPY.

From: Heineke, Merle J
Sent: Tuesday, May 7, 2019 2:27 PM
To: Lele, Aaditi <aaditi.lele@wsu.edu>; Liu, David <d.liu@wsu.edu>
Cc: Keeton, Denise <ddkeeton@wsu.edu>; Woodford, Nina Lynne <nwoodford@wsu.edu>; ocv.animal.shipping <ocv.animal.shipping@wsu.edu>
Subject: RE: Mice marked for shipment to UMASS

Dr. Woodford is aware of this. The requested health reports have been sent to UMASS and they have approved the shipment.

Thanks,
MErle

From: Lele, Aaditi <aaditi.lele@wsu.edu>
Sent: Tuesday, May 7, 2019 2:17 PM
To: Heineke, Merle J <merle.heineke@wsu.edu>; Liu, David <d.liu@wsu.edu>
Cc: Keeton, Denise <ddkeeton@wsu.edu>; Woodford, Nina Lynne <nwoodford@wsu.edu>; ocv.animal.shipping

<ocv.animal.shipping@wsu.edu>

Subject: RE: Mice marked for shipment to UMASS

Hi Merle and Dr. Liu

Since, there is no difference in the material, we don't need an MTA for the transfer. However, Dr. Liu needs to follow the animal research compliance regulations in order to have the mice shipped back. I have copied Nina Woodford, who can guide you about the necessary paperwork.

Thank you !

Regards,
Aaditi

From: Liu, David

Sent: Tuesday, May 7, 2019 11:53 AM

To: Lele, Aaditi <aaditi.lele@wsu.edu>; Heineke, Merle J <merle.heineke@wsu.edu>

Cc: Keeton, Denise <ddkeeton@wsu.edu>

Subject: RE: Mice marked for shipment to UMASS

No difference from what I received from Dr. Green.

Not to Cornell. UMASS is the only place.

From: Lele, Aaditi

Sent: Tuesday, May 07, 2019 11:02 AM

To: Heineke, Merle J <merle.heineke@wsu.edu>; Liu, David <d.liu@wsu.edu>

Cc: Keeton, Denise <ddkeeton@wsu.edu>

Subject: RE: Mice marked for shipment to UMASS

Hi Merle and Dr. Liu,

Is there any difference between the mice that you are returning now (as in any genetic modifications etc.) and the mice that you got originally from Dr. Green?

Also, if I have not misunderstood, you are planning to send some mice strains to Cornell too right? If that's the case, I'll need a filled out checklist for that as well so that I can draft the MTA with Cornell. Let me know.

Thanks !

Regards,
Aaditi

From: Heineke, Merle J

Sent: Tuesday, May 7, 2019 10:00 AM

To: Lele, Aaditi <aaditi.lele@wsu.edu>

Subject: RE: Mice marked for shipment to UMASS

Aaditi,

Please note that these animals originally came from Dr. Green at UMASS. Dr. Green is asking that the remaining mice be returned to him since Dr. Liu is leaving WSU. Is another MTA needed?

Merle

From: Lele, Aaditi <aaditi.lele@wsu.edu>

Sent: Tuesday, May 7, 2019 9:49 AM

To: Liu, David <d.liu@wsu.edu>; Heineke, Merle J <merle.heineke@wsu.edu>

Cc: Keeton, Denise <ddkeeton@wsu.edu>; Woodford, Nina Lynne <nwoodford@wsu.edu>

Subject: RE: Mice marked for shipment to UMASS

Dr. Liu,

Thank you for completing the checklist! I'll work on this and let you know if I need anything else.

Thanks !

Regards,

Aaditi

From: Liu, David

Sent: Tuesday, May 7, 2019 9:31 AM

To: Lele, Aaditi <aaditi.lele@wsu.edu>; Heineke, Merle J <merle.heineke@wsu.edu>

Cc: Keeton, Denise <ddkeeton@wsu.edu>; Woodford, Nina Lynne <nwoodford@wsu.edu>

Subject: RE: Mice marked for shipment to UMASS

Good morning Aaditi,

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Thank you.

David

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Subject: RE: Mice marked for shipment to UMASS

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We have received your MTA request. Please fill out the checklist attached with this e-mail and send it back to me. I'll begin drafting the MTA based on the information. Please let me know if you have any questions.

Thank you !

Regards,

Aaditi

Aaditi Lele

Technology Licensing Assistant

Washington State University

Lighty Student Services 280/286

Pullman, WA 99164-1060

Phone: 509-339-4363

aaditi.lele@wsu.edu



From: Liu, David <d.liu@wsu.edu>
Sent: Monday, May 6, 2019 5:03 PM
To: Simone, Tessa <Tessa.Simone@umassmed.edu>
Cc: Green, Michael <Michael.Green@umassmed.edu>; commercialization@wsu.edu; Heineke, Merle J <merle.heineke@wsu.edu>
Subject: Re: Mice marked for shipment to UMASS

Hi Merle,

I contacted the Office of Commercialization, as your suggested, but had not got any response yet and wondered if the email address is the correct one. Can you help to make sure that the OC had received my request together with the two forms I sent to them last week, and see if they can process the forms ASAP. As Tessa of UMASS mentioned in her email (see below), the forms need to be returned to UMASS as the first step for the transfer.

Thank you.

David

Sent from my Samsung Galaxy smartphone.

----- Original message -----

From: "Simone, Tessa" <Tessa.Simone@umassmed.edu>
Date: 5/6/19 2:03 PM (GMT-08:00)
To: "Liu, David" <d.liu@wsu.edu>
Cc: "Green, Michael" <Michael.Green@umassmed.edu>
Subject: Re: Mice marked for shipment to UMASS

Hi David,

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Tessa

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To: Lele, Aaditi <aaditi.lele@wsu.edu>; Heineke, Merle J <merle.heineke@wsu.edu>
Cc: Keeton, Denise <ddkeeton@wsu.edu>
Subject: RE: Mice marked for shipment to UMASS

No difference from what I received from Dr. Green.
Not to Cornell. UMASS is the only place.

From: Lele, Aaditi
Sent: Tuesday, May 07, 2019 11:02 AM
To: Heineke, Merle J <merle.heineke@wsu.edu>; Liu, David <d.liu@wsu.edu>
Cc: Keeton, Denise <ddkeeton@wsu.edu>
Subject: RE: Mice marked for shipment to UMASS

Hi Merle and Dr. Liu,

Is there any difference between the mice that you are returning now (as in any genetic modifications etc.) and the mice that you got originally from Dr. Green?

Also, if I have not misunderstood, you are planning to send some mice strains to Cornell too right? If that's the case, I'll need a filled out checklist for that as well so that I can draft the MTA with Cornell. Let me know.

Thanks !

Regards,
Aaditi

From: Heineke, Merle J
Sent: Tuesday, May 7, 2019 10:00 AM
To: Lele, Aaditi <aaditi.lele@wsu.edu>
Subject: RE: Mice marked for shipment to UMASS

Aaditi,
Please note that these animals originally came from Dr. Green at UMASS. Dr. Green is asking that the remaining mice be returned to him since Dr. Liu is leaving WSU. Is another MTA needed?
Merle

From: Lele, Aaditi <aaditi.lele@wsu.edu>
Sent: Tuesday, May 7, 2019 9:49 AM
To: Liu, David <d.liu@wsu.edu>; Heineke, Merle J <merle.heineke@wsu.edu>
Cc: Keeton, Denise <ddkeeton@wsu.edu>; Woodford, Nina Lynne <nwoodford@wsu.edu>
Subject: RE: Mice marked for shipment to UMASS

Dr. Liu,
Thank you for completing the checklist! I'll work on this and let you know if I need anything else.
Thanks !

Regards,
Aaditi

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To: Lele, Aaditi <aaditi.lele@wsu.edu>; Heineke, Merle J <merle.heineke@wsu.edu>
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Subject: RE: Mice marked for shipment to UMASS

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Thank you.
David

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Cc: Keeton, Denise <ddkeeton@wsu.edu>; Woodford, Nina Lynne <nwoodford@wsu.edu>
Subject: RE: Mice marked for shipment to UMASS

Hello Dr. Liu,
We have received your MTA request. Please fill out the checklist attached with this e-mail and send it back to me. I'll begin drafting the MTA based on the information. Please let me know if you have any questions.
Thank you !

Regards,
Aaditi

Aaditi Lele
Technology Licensing Assistant

Washington State University
Lighty Student Services 280/286
Pullman, WA 99164-1060
Phone: 509-339-4363
aaditi.lele@wsu.edu



From: Liu, David <d.liu@wsu.edu>
Sent: Monday, May 6, 2019 5:03 PM
To: Simone, Tessa <Tessa.Simone@umassmed.edu>
Cc: Green, Michael <Michael.Green@umassmed.edu>; commercialization@wsu.edu; Heineke, Merle J <merle.heineke@wsu.edu>
Subject: Re: Mice marked for shipment to UMASS

Hi Merle,

I contacted the Office of Commercialization, as your suggested, but had not got any response yet and wondered if the email address is the correct one. Can you help to make sure that the OC had received my request together with the two forms I sent to them last week, and see if they can process the forms ASAP. As Tessa of UMASS mentioned in her email (see below), the forms need to be returned to UMASS as the first step for the transfer.

Thank you.
David

Sent from my Samsung Galaxy smartphone.

----- Original message -----

From: "Simone, Tessa" <Tessa.Simone@umassmed.edu>
Date: 5/6/19 2:03 PM (GMT-08:00)
To: "Liu, David" <d.liu@wsu.edu>
Cc: "Green, Michael" <Michael.Green@umassmed.edu>
Subject: Re: Mice marked for shipment to UMASS

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Tessa

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Please help to make the shipment ASAP. Let me know if you have any questions.

Thank you.

David

From: Woodford, Nina Lynne
Sent: Tuesday, May 07, 2019 3:28 PM
To: Keeton, Denise; Heineke, Merle J; Lele, Aaditi; Liu, David
Cc: ocv.animal.shipping
Subject: RE: Mice marked for shipment to UMASS

If the MTA is not necessary and UMASS has accepted the health report then the shipment is approved to go. Please proceed with arranging the shipment

Nina Woodford DVM, MPH DACLAM
Director and Attending Veterinarian
Office of the Campus Veterinarian
Washington State University
509-335-6246
nwoodford@wsu.edu

From: Keeton, Denise <ddkeeton@wsu.edu>
Sent: Tuesday, May 7, 2019 2:50 PM
To: Heineke, Merle J <merle.heineke@wsu.edu>; Lele, Aaditi <aaditi.lele@wsu.edu>; Liu, David <d.liu@wsu.edu>
Cc: Woodford, Nina Lynne <nwoodford@wsu.edu>; ocv.animal.shipping <ocv.animal.shipping@wsu.edu>
Subject: RE: Mice marked for shipment to UMASS

Nina,
There is no further paperwork needed on our end. These mice originated from UMASS with the same PI they are going back to. No MTA to send them back.

Thanks,
Denise

Denise Keeton
Assistant Director, Business Operations
Office of Commercialization
Washington State University
Lighty 280/286
Pullman WA 99164-1060
(o) 509-335-1210 (f) 509-335-7237
ddkeeton@wsu.edu

Go Cougs!

THE INFORMATION CONTAINED IN THIS TRANSMISSION IS CONFIDENTIAL, MAY BE LEGALLY PRIVILEGED, AND IS ONLY FOR THE INTENDED RECIPIENT IDENTIFIED ABOVE. IF YOU HAVE RECEIVED THIS TRANSMISSION IN ERROR, PLEASE IMMEDIATELY NOTIFY US BY TELEPHONE, RETURN THE ORIGINAL MESSAGE TO US, AND RETAIN NO COPY.

From: Heineke, Merle J
Sent: Tuesday, May 7, 2019 2:27 PM
To: Lele, Aaditi <aaditi.lele@wsu.edu>; Liu, David <d.liu@wsu.edu>
Cc: Keeton, Denise <ddkeeton@wsu.edu>; Woodford, Nina Lynne <nwoodford@wsu.edu>; ocv.animal.shipping <ocv.animal.shipping@wsu.edu>
Subject: RE: Mice marked for shipment to UMASS

Dr. Woodford is aware of this. The requested health reports have been sent to UMASS and they have approved the shipment.

Thanks,
MErle

From: Lele, Aaditi <aaditi.lele@wsu.edu>
Sent: Tuesday, May 7, 2019 2:17 PM
To: Heineke, Merle J <merle.heineke@wsu.edu>; Liu, David <d.liu@wsu.edu>
Cc: Keeton, Denise <ddkeeton@wsu.edu>; Woodford, Nina Lynne <nwoodford@wsu.edu>; ocv.animal.shipping <ocv.animal.shipping@wsu.edu>
Subject: RE: Mice marked for shipment to UMASS

Hi Merle and Dr. Liu

Since, there is no difference in the material, we don't need an MTA for the transfer. However, Dr. Liu needs to follow the animal research compliance regulations in order to have the mice shipped back. I have copied Nina Woodford, who can guide you about the necessary paperwork.

Thank you !

Regards,
Aaditi

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Aaditi

Aaditi Lele

Technology Licensing Assistant

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Lighty Student Services 280/286

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Phone: 509-339-4363

aaditi.lele@wsu.edu



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Please help to make the shipment ASAP. Let me know if you have any questions.

Thank you.

David

From: Durfee, Codie Jo
Sent: Friday, August 16, 2019 8:58 AM
To: White, Stephen; or.ocv.alert; Luft, Jan Elizabeth; Libby, Julia E
Cc: [REDACTED] 38; Durfee, Codie; 'Stephen.White@wsu.edu'; Fry, Lindsay Michelle
Subject: RE: Mouse update RE: Culling mice - request training

Whoops! I was just a smidge off on my count once I put everything in a spreadsheet. There are actually 59 mice (3 cages have neonates).

Sorry for the confusion.
Codie

From: Durfee, Codie Jo
Sent: Thursday, August 15, 2019 2:48 PM
To: White, Stephen <stephen.white@usda.gov>; or.ocv.alert <or.ocv.alert@wsu.edu>; Luft, Jan Elizabeth <jluft@wsu.edu>; Libby, Julia E <jlibby@wsu.edu>
Cc: [REDACTED] 38 Durfee, Codie <codie.durfee@usda.gov>; 'Stephen.White@wsu.edu' <Stephen.White@wsu.edu>; Fry, Lindsay Michelle <lfry@wsu.edu>
Subject: Mouse update RE: Culling mice - request training

Hello Everyone!

Just a quick little update.

Levi is coming by to update our signage tomorrow at 2:30 for the experimental room. I will meet him downstairs to find out what else we might need for our room. Jan might join us.

I went through training today with Johnathan DenHerder on culling mice. He taught me how to do this for adults and the neonates/pinkies. It took me about 3 hours to get through all the mice we had planned for and took a new inventory. I went ahead and did all we planned for excluding 4 females. Johnathan said it would be good to hold a few back for [REDACTED] 33 for when he is back from his trip to practice dissection and learn how to euthanize them. I think it would be good for both of us to attend the dissection training to save some time.

We now have 21 cages with 56 mice (3 of these have pups). I plan to update our excel sheet tomorrow, but I have run out of time for today. All seem to be in good health. One male in cage 3 that has been under watch for barbering was seen scratching his barbered left cheek and side. I am going to monitor him to make sure he doesn't have any sores. So far his skin looks healthy aside from being bald. He is in a cage with fairly new pups so I don't want to disturb them too much.

Please let me know if you have any questions.

Thank you,
Codie

From: White, Stephen [<mailto:stephen.white@usda.gov>]
Sent: Tuesday, August 13, 2019 4:40 PM
To: or.ocv.alert <or.ocv.alert@wsu.edu>; Luft, Jan Elizabeth <jluft@wsu.edu>; Libby, Julia E <jlibby@wsu.edu>
Cc: [REDACTED] 38 Durfee, Codie <codie.durfee@usda.gov>; Durfee, Codie Jo

<chanke@wsu.edu>; 'Stephen.White@wsu.edu' <Stephen.White@wsu.edu>; Fry, Lindsay Michelle <lfry@wsu.edu>

Subject: Culling mice - request training

Hi all,

Thanks for your help as we have established and begun learning about our new mouse colonies. After a bit more success than we had feared initially, we need to resize our breeding colony. In total we plan to cull 11 adults, 2 unweaned litters, and 37 weaned pups. Our protocol for euthanasia is by CO2 with either cervical dislocation, exsanguination, or decapitation as a secondary method. Since we do not need blood, cervical dislocation would be our preferred secondary method for its reliability and expedience.

We have not euthanized a mouse in our lab before and would need training. As such, we would like to coordinate with the vivarium and/or OCV to be shown the euthanasia of these mice as training. [REDACTED] 38 will be out of the office August 15 through August 26 but Codie Durfee will be here during that time.

Below is a table of which cages we plan to euthanize vs. keep in the breeding colony. The table is organized by age (adult vs. weaned pup) and genotype (TLR2 knockout vs. non-knockout Black 6 wild-type mice). This is also attached as a Word document.

What should be done to work out training?

Thank you!

Stephen

<u>Cull</u>	<u>Keep</u>
<u>TLR2 knockout adults</u> Cage 1: 2 adults (male, female pair) + pups Cage 4: 1 adult (male) Cage 5: 2 adults (male, female pair)	<u>TLR 2 knockout adults</u> Cage 2: 2 adults (male, female pair) + pups Cage 3: 2 adults (male, female pair) + pups
<u>Black 6 (wild-type) adults</u> Cage 6: 2 adults (male, female pair) Cage 8: 2 adults (male, female pair) + pups Cage 9: 2 adults (male, female pair)	<u>Black 6 (wild-type) adults</u> Cage 7: 2 adults (male, female pair) + pups Cage 10: 2 adults (male, female pair) + pups
<u>TLR2 knockout weaned pups</u> Cage 11: 4 male pups Cage 12: 2 female pups Cage 13: 3 male pups Cage 14: 5 female pups Cage 15: 4 female pups Cage 21: 3 male pups Cage 23: 3 male pups Cage 24: 3 female pups Cage 25: 2 male pups	<u>TLR2 knockout weaned pups</u> Cage 20: 3 male pups Cage 22: 3 female pups Cage 29: 2 male pups Cage 30: 2 male pups
<u>Black 6 (wild-type) weaned pups</u> Cage 16: cull 2 pups (male), leave other 3 Cage 17: 1 female pup	<u>Black 6 (wild-type) weaned pups</u> Cage 16: keep 3 pups (male), cull other 2 Cage 19: 3 female pups Cage 26: 4 male pups Cage 27: 3 female pups Cage 28: 3 female pups

Cage 18: 5 male pups	
<u>Total adults culled:</u> 11 + 2 groups unweaned pups <u>Total weaned pups culled:</u> 37	<u>Total adults kept:</u> 8 + 4 groups unweaned pups <u>Total weaned pups kept:</u> 27

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From: Robbins, Charles T
Sent: Thursday, June 06, 2019 3:56 PM
To: Woodford, Nina Lynne; or.ocv.alert; Evans Hutzenbiler, Brandon Dean; Jansen, Heiko; Blatner, Keith Allan
Cc: Ekstrand, Alan
Subject: RE: New Guides Offer Helpful Information on Caring for Animals

All: This USDA document was prepared without much scientific knowledge. Examples:

1. "allowing bears to eat as much as they wish in the fall"---that is a recipe for immense obesity.
2. "Captive bears do not truly hibernate." This was a popular argument 30 Or 40 yrs ago when people debated whether bears entered dormancy vs hibernation. That discussion is pointless and not helpful.
3. "---bears ---require food during hibernation"---Absolutely not true. There would be millions of dead bears in the wild if this were true. This and point 1 are designed to produce morbidly obese bears.

Charlie

From: Woodford, Nina Lynne
Sent: Thursday, June 06, 2019 11:00 AM
To: or.ocv.alert <or.ocv.alert@wsu.edu>; Evans Hutzenbiler, Brandon Dean <brandon.hutzenbiler@wsu.edu>; Robbins, Charles T <ctrobbins@wsu.edu>; Jansen, Heiko <heiko@wsu.edu>; Blatner, Keith Allan <blatner@wsu.edu>
Cc: Ekstrand, Alan <alan.ekstrand@wsu.edu>
Subject: FW: New Guides Offer Helpful Information on Caring for Animals

Passing this on as USDA now has basic informational guidance on bear care. Not a new regulatory document and it appears this is targeted at exotic pet owners/small exhibitors/zoos and not research facilities. Looking it over, we follow the guidance for the most part but I see some differences on the "bear nutrition & winter lethargy" page.

https://www.aphis.usda.gov/aphis/ourfocus/animalwelfare/caw/at_caw_animal_care_aids

We should be familiar with the documents and be able to justify where we differ in practices if asked during a visit (i.e. we study nutrition and hibernation). Again these are not new regulations

Let me know if you have questions

Thanks

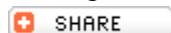
Nina

Nina Woodford DVM, MPH DACLAM
Director and Attending Veterinarian
Office of the Campus Veterinarian
Washington State University
509-335-6246
nwoodford@wsu.edu

From: APHIS Animal Care <ace@subscribers.usda.gov>
Sent: Thursday, June 6, 2019 10:38 AM
To: Woodford, Nina Lynne <nwoodford@wsu.edu>
Subject: New Guides Offer Helpful Information on Caring for Animals



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New Guides Offer Helpful Information on Caring for Animals

Animal Care is developing easy-to-understand informational guidance to help anyone who owns or cares for animals.

These one-page documents, called Animal Care Aids, cover a wide range of topics and species, including canine dental and coat care, primate environmental enhancement, bear care, and special environmental needs of semi-aquatic and flying mammals. Newly added Animal Care Aids include: Feeding Nondomestic Cats, Metabolic Bone Disease in Nondomestic Cats, and Tips and Calculations: Adding Calcium to Boneless Meat Diets for Nondomestic Cats. The Animal Care Aids are posted [here](#).

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From: Woodford, Nina Lynne
Sent: Friday, June 07, 2019 7:46 AM
To: Robbins, Charles T; or.ocv.alert; Evans Hutzenbiler, Brandon Dean; Jansen, Heiko; Blatner, Keith Allan
Cc: Ekstrand, Alan
Subject: RE: New Guides Offer Helpful Information on Caring for Animals

I also took note of those points when I read it. We have ample scientific justification to counter if questioned but wanted you to be prepared. Again these are not new regulation so I don't know if these guides will ever come up for discussion on a site visit (but they could)

Nina Woodford DVM, MPH DACLAM
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From: Cheng, Zhaokang
Sent: Tuesday, June 18, 2019 11:12 AM
To: Packer, Katherine
Cc: or.ocv.alert; Clyde, Gaylynn Goolsby; Chen, Jingrui
Subject: Re: Packing Slip Needed: Cheng - Buprenorphine SR-LAB 5ml (0.5mg/ml)

Thank you Kay. I am copying Jingrui on this email, so he will be able to email the invoice to you as instructed.
ZK

From: Packer, Katherine
Sent: Tuesday, June 18, 2019 9:25 AM
To: Cheng, Zhaokang
Cc: or.ocv.alert; Clyde, Gaylynn Goolsby
Subject: Packing Slip Needed: Cheng - Buprenorphine SR-LAB 5ml (0.5mg/ml)

Hello,

Your Buprenorphine SR order should arrive to you between 06-20-2019 and 06-21-2019.

When your order arrives:

1. Inspect to make sure all items arrived
2. Write on the invoice the date received and sign your name
3. Scan/email a copy of the invoice to karp.packer@wsu.edu

If you have any questions, please feel free to contact me.

Thank you,

Kay

Katherine Packer, Fiscal Specialist 2

Office of the Campus Veterinarian
Washington State University
PO Box 641165 (100 Dairy Road)
Pullman, WA 99164-1165
Office Phone: (509) 335-4230
Main Line: (509) 335-6246
Fax: (509) 335-3162
Email: karp.packer@wsu.edu



From: Clyde, Gaylynn Goolsby <gclyde@wsu.edu>
Sent: Monday, June 17, 2019 3:07 PM
To: Cheng, Zhaokang <zhaokang.cheng@wsu.edu>
Cc: or.ocv.alert <or.ocv.alert@wsu.edu>
Subject: RE: Buprenorphine SR-LAB 5ml (0.5mg/ml)

So sorry for the delay. It was supposed to be ordered on Friday, but it appears it was not. We will get it ordered first thing in the morning and hopefully it will arrive to you all on Friday in Spokane.

Gay Lynn Clyde, DVM
Assistant Director Campus Veterinary Services
Office of Campus Veterinarian
Washington State University
Pullman, WA. 99164-1165
Phone (509) 335-4991 or (509) 335-6246
Fax (509) 335-3162
Email: gclyde@wsu.edu
Program URL: <http://campusvet.wsu.edu>

From: Cheng, Zhaokang <zhaokang.cheng@wsu.edu>
Sent: Monday, June 17, 2019 2:48 PM
To: Clyde, Gaylynn Goolsby <gclyde@wsu.edu>
Subject: Re: Buprenorphine SR-LAB 5ml (0.5mg/ml)

Hi Dr. Clyde,

I haven't heard from you about my previous email request. Could you please let me know how to order Bup-SR?

Thanks,

ZK

From: Cheng, Zhaokang

Sent: Thursday, June 13, 2019 1:34:57 PM

To: Clyde, Gaylynn Goolsby

Subject: Buprenorphine SR-LAB 5ml (0.5mg/ml)

Hi Dr. Clyde,

Could you please order 1 vial of Buprenorphine SR-LAB 5ml (0.5mg/ml) for use in mice (ASAF#6480)? Thank you,

ZK

From: Packer, Katherine
Sent: Tuesday, June 18, 2019 11:41 AM
To: Cheng, Zhaokang
Cc: or.ocv.alert; Clyde, Gaylynn Goolsby; Chen, Jingrui
Subject: RE: Packing Slip Needed: Cheng - Buprenorphine SR-LAB 5ml (0.5mg/ml)

Thank you for your help,

Kay ☺

Katherine Packer, Fiscal Specialist 2

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Fax: (509) 335-3162
Email: karp.packer@wsu.edu



From: Cheng, Zhaokang <zhaokang.cheng@wsu.edu>
Sent: Tuesday, June 18, 2019 11:12 AM
To: Packer, Katherine <karp.packer@wsu.edu>
Cc: or.ocv.alert <or.ocv.alert@wsu.edu>; Clyde, Gaylynn Goolsby <gclyde@wsu.edu>; Chen, Jingrui <jingrui.chen@wsu.edu>
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ZK

From: Clyde, Gaylynn Goolsby
Sent: Friday, September 06, 2019 3:51 PM
To: Matuk Sarinana, Celina Martiza
Cc: or.ocv.alert
Subject: RE: Requesting brand inspection

I will head out that way right now. I will leave the paper work on your desk.

From: Matuk Sarinana, Celina Martiza <c.matuksarinana@wsu.edu>
Sent: Friday, September 6, 2019 3:40 PM
To: Woodford, Nina Lynne <nwoodford@wsu.edu>; Clyde, Gaylynn Goolsby <gclyde@wsu.edu>
Subject: Requesting brand inspection

Greetings,

We have three bull calves for sale at the dairy and a customer willing to take them tomorrow when he passes through Pullman to Bonners Ferry Idaho.

Is by any chance possible to have them brand inspected today?

Sorry for the very late notice but I just received the request. I will confirm the purchaser later if we are going to be able to sell those to him tomorrow.

Purchaser:



Thank you

Celina

Celina M. Matuk-Sarinana, M.S. DVM
Dairy Farms Operation Manager
Washington State University

From: Matuk Sarinana, Celina Martiza
Sent: Friday, September 06, 2019 4:12 PM
To: Clyde, Gaylynn Goolsby
Cc: or.ocv.alert
Subject: Re: Requesting brand inspection

Thank you!

Celina M. Matuk-Sarinana, M.S. DVM
Dairy Farms Operation Manager
Washington State University

From: Clyde, Gaylynn Goolsby <gclyde@wsu.edu>
Sent: Friday, September 6, 2019 3:50:44 PM
To: Matuk Sarinana, Celina Martiza <c.matuksarinana@wsu.edu>
Cc: or.ocv.alert <or.ocv.alert@wsu.edu>
Subject: RE: Requesting brand inspection

I will head out that way right now. I will leave the paper work on your desk.

From: Matuk Sarinana, Celina Martiza <c.matuksarinana@wsu.edu>
Sent: Friday, September 6, 2019 3:40 PM
To: Woodford, Nina Lynne <nwoodford@wsu.edu>; Clyde, Gaylynn Goolsby <gclyde@wsu.edu>
Subject: Requesting brand inspection

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Purchaser:



Thank you

Celina

Celina M. Matuk-Sarinana, M.S. DVM
Dairy Farms Operation Manager
Washington State University

From: Clyde, Gaylynn Goolsby
Sent: Tuesday, July 02, 2019 5:08 PM
To: Matuk Sarinana, Celina Martiza; Woodford, Nina Lynne
Cc: or.ocv.alert
Subject: RE: Requesting brand inspection

Hi Celina,

I can be out in the afternoon on Wednesday around 2pm.

Thank you,

Gay Lynn Clyde, DVM
Assistant Director Campus Veterinary Services
Office of Campus Veterinarian
Washington State University
Pullman, WA. 99164-1165
Phone (509) 335-4991 or (509) 335-6246
Fax (509) 335-3162
Email: gclyde@wsu.edu
Program URL: <http://campusvet.wsu.edu>

From: Matuk Sarinana, Celina Martiza <c.matuksarinana@wsu.edu>
Sent: Tuesday, July 2, 2019 11:34 AM
To: Clyde, Gaylynn Goolsby <gclyde@wsu.edu>; Woodford, Nina Lynne <nwoodford@wsu.edu>
Subject: Requesting brand inspection

Dr. Woodford and Dr. Clyde,

I hope that this email find you well. Currently we have eight bulls calves at the dairy. I have somebody interested in two calves from Idaho. Would it be possible to have them brand inspected by Wednesday? Friday should also work as well since the calves are getting picked up by 6 pm on that day.

Purchaser:



Thank you for your time and help.

Celina Matuk

Celina M. Matuk-Sarinana, M.S. DVM
Dairy Farms Operation Manager
Washington State University

From: McCleary, Jessie May
Sent: Thursday, August 29, 2019 9:56 AM
To: Appleyard, Suzanne; Zhu, Mingyan
Cc: or.ocv.alert
Subject: RE: Required IACUC Training: AST-O

Hi Suzy, hope all your travels went well. Could you remind me who all needs the training?
It is probably easiest for you to look at the training schedule and pick a date that would work: <https://iacuc.wsu.edu/hands-on-training/>
Let me know if there are any questions.

Thank you,
Jess

From: Appleyard, Suzanne <s.appleyard@wsu.edu>
Sent: Thursday, August 29, 2019 9:51 AM
To: McCleary, Jessie May <jmccleary@wsu.edu>; Zhu, Mingyan <mzhu115@wsu.edu>
Subject: Re: Required IACUC Training: AST-O

Hi Jessie,

I was wondering whether we could schedule a hands on training for people in my lab who need an updated training session. As we were traveling extensively this summer Ashley kindly said we could do it when we returned as long as we weren't performing surgeries. As the new semester is up and going we would like to take this training. What times would be available for the next month?

I hope you had a good summer.

Thank you in advance for your help.

Suzy

From: McCleary, Jessie May <jmccleary@wsu.edu>
Sent: Wednesday, May 29, 2019 8:53 AM
To: Appleyard, Suzanne <s.appleyard@wsu.edu>; Zhu, Mingyan <mzhu115@wsu.edu>
Subject: RE: Required IACUC Training: AST-O

Suzy, that would work great. All trainings in June are currently available. Let me know which one works for you. As people request trainings, the website will be updated to show what is scheduled.

Let me know if you have questions,
Jess

From: Appleyard, Suzanne <s.appleyard@wsu.edu>
Sent: Wednesday, May 29, 2019 7:48 AM

To: Zhu, Mingyan <mzhu115@wsu.edu>; McCleary, Jessie May <jmccleary@wsu.edu>
Subject: Re: Required IACUC Training: AST-O

Hi Jessie,
As several of my lab need the training we can try to coordinate a day that would work for you and them.
Thank you for your help.
Suzy

From: Zhu, Mingyan
Sent: Wednesday, May 29, 2019 6:43:24 AM
To: McCleary, Jessie May
Cc: Appleyard, Suzanne
Subject: RE: Required IACUC Training: AST-O

Hi Jessie,

Thank you for information. I believe that I've finished that workshop 10 years ago in same place.
Can I meet you today (5/29/2019) 10-12 am to join this workshop?
I think ASAF number is 6528 for protocol number 3682 under

Thanks.

Mingyan

From: McCleary, Jessie May
Sent: Tuesday, May 28, 2019 2:20 PM
To: Zhu, Mingyan <mzhu115@wsu.edu>
Cc: Appleyard, Suzanne <s.appleyard@wsu.edu>; or.ocv.alert <or.ocv.alert@wsu.edu>
Subject: RE: Required IACUC Training: AST-O

Hello Dr Zhu,
I am a LVT from the Office of the Campus Veterinarian. Please let me know when you have completed the completed the AST-O training and we can get you scheduled for the hands-on portion of the training (AST-H).
Here is the link to the days available in June: <https://iacuc.wsu.edu/hands-on-training/>

[OCV Hands on training | Institutional Animal Care and Use Committee | Washington State University - iacuc.wsu.edu](https://iacuc.wsu.edu/hands-on-training/)

iacuc.wsu.edu

Basic Mouse Handling and Rat Handling includes: Use of personal protective equipment, handling and restraint of rodents, giving injections, collecting blood, euthanasia techniques, plus any specific procedures needed for your laboratory protocol. Aseptic Surgical Technique- hands on workshop includes: Training in aseptic techniques to minimize post-operative infections, instrument ...

Could you please send me the ASAF number for the protocol you are working under?

I have attached the outline of the training for you to look over and a map to our building. Let me know if you have any questions. I am looking forward to meeting with you.

Thank you,

Jessie McCleary, LVT
Washington State University
Office of the Campus Veterinarian
jmccleary@wsu.edu
Office (509)335-2595
Main line (509)335-6246
Emergency phone (509)330-1871

From: IACUC <or.ora.iacuc@wsu.edu>

Sent: Friday, May 17, 2019 9:05 AM

To: Zhu, Mingyan <mzhu115@wsu.edu>

Cc: Appleyard, Suzanne <s.appleyard@wsu.edu>; or.ocv.alert <or.ocv.alert@wsu.edu>

Subject: Required IACUC Training: AST-O

Importance: High

Hi Dr. Zhu,

This is a courtesy notice that you are required for ASAF #6528 to complete the ***Aseptic Surgical Technique WSU (AST-O)***, an online course on AALAS Learning Library website that is required by WSU IACUC for all new researchers/ faculty/ staff performing surgery on research or teaching animals. Board-certified surgeons working within their specialty may be exempt.

Personnel must complete training before ASAF #6528 can be approved.

Please note: As of May 4th, 2018 per the WSU IACUC Policy #20, all required online training must be completed prior to approval.

****All Principal Investigators need to submit an amendment prior to the approval of personnel working on the protocol. For the new online system instructions, please visit <https://myresearch.wsu.edu/Compliance/IACUC/ASAFInstructions.aspx>. For the old PDF system form, please visit <https://iacuc.wsu.edu/forms/>.**

Our records indicate that you already have an account with AALAS Learning Library.

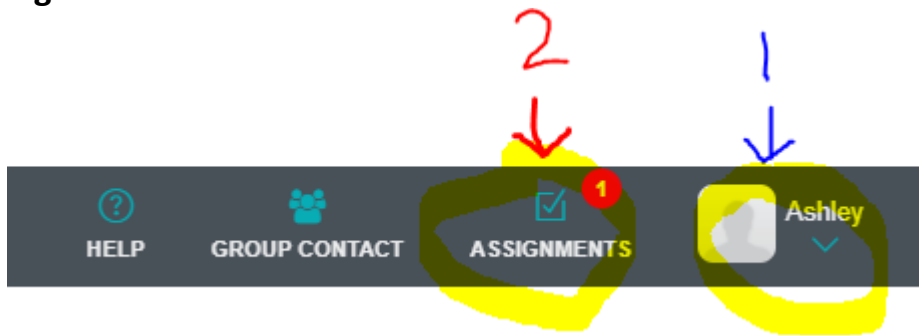
- The link to log in is: <https://aalaslearninglibrary.org/index.html#/login/signin>
- Enter your User name: 41
- Enter your password. If you need to reset your password, you can click on the “I forgot my password” link. If that does not work, contact the Animal Welfare Program.

Step 1: Once you log on, you will be directed to the AALAS homepage. Complete your profile located in the upper right corner with the person icon (arrow 1 in Figure 1).

Step 2: After you complete all required fields, find the **Aseptic Surgical Technique WSU (AST-O)** under the “Assignments” tab located in the top right (arrow 2 in Figure 1). The course and exam together can take approximately 1 hour.

Once you have passed the exam, you have completed the course.

Figure 1:



*If the exam link does not appear, a lesson has been missed. Please go to your profile (blue arrow 1 in Figure 1), then click on My Transcript to see which lessons are remaining.

To set up hands-on training after the completion of all required online training, please contact the Office of the Campus Veterinarian at or.ocv.alert@wsu.edu or (509) 335-6246.
DVM, LVT, MD, and RN may be exempt from the hands-on course.

If you have any issues regarding IACUC training, please contact the Animal Welfare Program at (509) 335-5353 or email at iacuc@wsu.edu.

Carmen Haines

Animal Welfare Program

Washington State University | Office of Research Assurances

iacuc@wsu.edu | Phone: (509) 335-1763

<https://iacuc.wsu.edu>

From: McCleary, Jessie May
Sent: Monday, August 26, 2019 4:00 PM
To: IACUC; Dilworth, Kayla Ann
Cc: Ritter, W Sue; or.ocv.alert
Subject: RE: Required IACUC Training: AST-O
Attachments: AST-H Rodents Outline 2018.docx; OCV Location.pptx

Hello Kayla,

I am a LVT from the Office of the Campus Veterinarian. Please let me know when you have completed the completed the AST-O training and we can get you scheduled for the hands-on portion of the training (AST-H).

Here is the link to the days available in Aug/Sept: <https://iacuc.wsu.edu/hands-on-training/>

Could you please send me the ASAF number for the protocol you are working under?

I have attached the outline of the training for you to look over and a map to our building. Let me know if you have any questions. I am looking forward to meeting with you.

Thank you,

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Washington State University
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Emergency phone (509)330-1871

From: IACUC <or.ora.iacuc@wsu.edu>
Sent: Monday, August 26, 2019 3:56 PM
To: Dilworth, Kayla Ann <kayla.dilworth@wsu.edu>
Cc: Ritter, W Sue <sritter@wsu.edu>; or.ocv.alert <or.ocv.alert@wsu.edu>
Subject: Required IACUC Training: AST-O
Importance: High

Hi Kayla,

This is a courtesy notice that you are required for Dr. Ritter's to complete the ***Aseptic Surgical Technique WSU (AST-O)***, an online course on AALAS Learning Library website that is required by WSU IACUC for all new researchers/ faculty/ staff performing surgery on research or teaching animals. Board-certified surgeons working within their specialty may be exempt.

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Our records indicate that you do not have an account with AALAS Learning Library. Once you are enrolled, you will receive enrollment emails from AALAS with log in information and links. If you do not see it in your inbox, check the spam/junk folder. Please let the Animal Welfare Program know if you need password setup email resent.

Step 1: Once you log on, you will be directed to the AALAS homepage. Complete your profile located in the upper right corner with the person icon (arrow 1 in Figure 1).

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Carmen Haines

Animal Welfare Program

Washington State University | Office of Research Assurances

iacuc@wsu.edu | Phone: (509) 335-1763

<https://iacuc.wsu.edu>

Aseptic Surgical Technique for Rodents

Hands-on Workshop

Washington State University
Office of the Campus Veterinarian-Vet Services

1. Introduction

- a. Discuss similarities & differences between rodent and non-rodent surgery
 - i. Similar- Both require asepsis for anything touching the surgical area
 - ii. Differences are primarily due to the small size of the rodent
 - 1. Designated space vs. dedicated facility (room or suite)
 - a. Clean lab bench in non-traffic area
 - 2. Clean lab coat vs. sterile surgical gown
 - 3. Work alone vs. surgical team
 - a. Without assistant, asepsis requires careful planning

2. Procedure Area Preparation

- a. Designated for surgery currently, no traffic/ drafts, post signs “Aseptic Surgery in Progress”
- b. Surgical/anesthesia records and disinfected pen
- c. Disinfect surfaces. Remove unneeded items.
- d. Animal Prep area: clippers, scrub solution, eye ointment, disposable layer
- e. Surgical Area: heat source, light, glass bead sterilizer? Absorbent clean or sterile layer, sterile instrument pack, drape and suture, (wait to open until just before gloves on)
- f. Post-operative area: heat source on lowest setting, under half of recovery cage, white pad or paper towels (instead of bedding), analgesic and saline drawn into syringes(warm syringes with heating pad)

3. Instruments and Materials Preparation

- a. Autoclaving- steam heat (moist heat, physical method)
 - i. Metal instruments, some catheter tubing, saline, water
 - ii. Packaging
 - 1. Surgical packs- gauze, drape, instruments
 - 2. Envelopes- Visi-peel
 - 3. Indicator tape, indicator strips
 - 4. Monitor autoclave quarterly with biological indicator
 - iii. Date autoclaved materials – maximum 6 month expiration
 - iv. Pre-packaged sterile supplies- suture, blades (check expiration dates)
- b. Glass bead sterilizers- dry heat (modified aseptic technique “tips only”)
 - i. Start with autoclaved pack then use hot glass beads between animals
 - ii. Wipe instruments to remove blood & debris before use

Revised 9/21/18

Obtained by Rise for Animals.
Uploaded to Animal Research Laboratory Overview (ARLO) on 02/03/2021

- iii. Instrument tips in heat for 30-60 seconds (too long may warp)
 - iv. Cool before use to prevent burns to animal or surgeon
 - v. New autoclave pack every 5 animals.
 - vi. Only tips are re-sterilized. Position: tips away/ handles toward surgeon
 - vii. New surgery gloves for each animal (unless assistant prepped new animal)
 - viii. Tips only surgical technique (handles & hands are clean but not sterile)
- c. Chemical sterilants- liquid “cold sterilization”
- i. Liquid sterilants require 6-8 hours contact times to kill spores
 - ii. Liquid must contact all surfaces (fill tubing)
 - iii. Rinse chemicals off with sterile saline/H₂O before use
 - iv. Alcohol is a disinfectant but not a sterilant
- d. Chemical sterilants- gas “cold and dry sterilization”
- i. Ethylene Oxide
 - ii. Vaporized Hydrogen Peroxide

4. Animal Preparation

- a. Anesthesia
- i. Injectable or inhalation- assemble needed supplies (DEA regulations)
 - ii. Weigh each animal pre-operatively & write in surgery/anesthesia record (grams)
 - iii. Calculate dosages and draw up anesthesia, analgesia,
 - iv. Give subcutaneous saline before the surgery if possible
 - v. No food or water restriction for rodents
 - vi. Rodents are susceptible to dehydration, hypoglycemia, hypothermia
- b. Eye ointment
- i. Anesthetized rodents don't close eyes
 - ii. Prevent drying of corneas
 - iii. Use ophthalmic ointment (artificial tears)
- c. Monitor anesthetic depth (variation in strains and individual animals)
- i. Observe respiratory rate and depth (Count 15 seconds X 4)
 - ii. Color of mucus membranes, ears, tail, feet
 - iii. Toe pinch withdrawal reflex? Use fingers or hemostat
 - 1. If withdraw, too light for surgery but clipping is OK
 - 2. Intra-operative- Maintain sterility using hemostats- handles sterile
 - iv. If using injectable anesthetic (ketamine/xylazine) only boost with ketamine (1/3rd).
- d. Clip hair
- i. Blades #40 - maintain with spray cleaner/lubricant
 - ii. Razor or Depilatory Cream (Nair) may be used

- e. Scrub: can be done with exam gloves
 - i. Chlorhexidine (scrub or solution) then alcohol OR Povidone/iodine (scrub or solution) then alcohol
 - ii. Start at center and move out in spiral to clipped margins
 - iii. Triple scrub then may apply solution to dry on (not scrub- contains soap)
 - iv. Cotton swabs versus gauze sponges
 - v. Don't soak animal or get disinfectant in eyes

5. Surgeon Preparation

- a. Required
 - i. Remove earrings, necklaces, rings, watches, scarves, etc...
 - ii. Clean lab coat
 - 1. Free of debris, hair, or bedding to fall into surgical area
 - 2. Snug lab coat cuffs or tape around wide sleeves or roll up sleeves
 - iii. Face mask
 - 1. protect surgical field from bacteria (surgeon's exhaled breath)
 - 2. Mask is clean but not sterile (Do not touch with gloved hands)
 - iv. Surgical gloves – Wait until animal is prepped & ready to be draped
Working alone? Before gloving ask yourself: **LIST** (Light on, Instrument pack open, Supplies/suture, Toe pinch check anesthesia/ Time starting noted)
 - 1. Individually packaged latex or nitrile surgical gloves
 - a. **NOT** exam gloves even if cleaned and/or autoclaved
 - b. Do not re-sterilize & re-use surgical gloves
 - c. Surgical gloves- check expiration date
 - d. Sized in half sizes to fit (example: 6.0, 6.5, 7.0, 7.5)
 - 2. Wash hands with soap and warm water. Dry with clean towel.
 - 3. Aseptic gloving technique- demonstrate & practice
 - 4. If gloves are contaminated during surgery– get new gloves
 - 5. Put on new sterile gloves every 3 animals or if they become contaminated between surgeries.
 - v. **Review your ASAF** surgery section on “How aseptic technique is maintained”
You must adhere to what is written in the ASAF.
- b. Recommended
 - i. Hair cap/ bonnet recommended. Tie back long hair. Beard cover.
 - ii. An Assistant: the surgeon can't touch anything non-sterile once gloved. Surgery is more efficient, faster and less complicated if there is an assistant.
 - iii. **Review ASAF**, if cap for example, is listed then need to wear.

6. Draping

- a. Open sterile instrument and drape pack → put on sterile gloves → then may handle drape

- b. Place drape & lift drape to check toe pinch with hemostats without contaminating sterile gloves.
- c. Alternative: Press N' Seal plastic wrap –use will be demonstrated at workshop

7. Perform surgery

- a. Intraoperative monitoring of anesthetic depth- surgical record
- b. Maintain sterile gloves and sterile field
- c. Batch surgery? Working alone? Heat sterilize instrument tips prior to handling animal.

8. Recovery & post-operative care

- a. Post-operative analgesic and warm saline
- b. Wrap rodent in huck or paper towel to maintain body heat
- c. Rotate position every 10 minutes until sternal, semi awake
- d. Maintain heat source & keep separate until animal is moving about cage
- e. Check surgery rodents end of first day
- f. Monitor twice daily for first 3 days, then once daily for appetite, activity, grooming
- g. Check incision for redness, swelling, discharge, suture failure
- h. Water-soaked kibble or Gel pack in cage bottom advised
- i. Document observations, analgesics, and date of suture removal (~10 days)
- j. Analgesia discussion: do what is in approved protocol

9. Conclusion

- a. Success requires careful planning & preparation
- b. Difficult to do surgery alone especially on multiple animals- recommend having assistant to help prepare and recover animals while surgeon focuses on surgery

OCV is located in the LARC Building (in the yellow circle), the orange star is where the entrance door to the building. Front office is on your left. Josh our receptionist will bring you to Training Room. If he is not at his desk ring the bell on the wall for assistance. (The blue star is the training room #198.)

