## Column E Explanation

This form is intended as an aid to complete the Column E explanation. It is not an official form and its use is voluntary. Annual Reports and explanations should NOT include PII information such as names (principle investigators and research staff), addresses, protocols, meeting notes (either in part of in full), the animals room numbers, grant information, veterinary care programs, and the like. A Column E explanation must be written so as to be understood by lay person as well as scientists.

Registration Number: 63-R-0107

2.	Number of animals used in this study.			
3.	Species (common name) Peromyscus of animals used in this study.			
4.	Explain the procedure producing pain and/or distress. Explanations should include a brief description of the procedure, but also explain what the animal's experience, examples of which may include, but are not limited to: Neurological signs, seizures, tremors, paralysis, lethargy, inappetance, respiratory signs, GI distress, vomiting, and diarrhea.			
Peromyscus will be infected with Borelia burgdorferi via subcutaneous injection. Blood is collected, under isoflurane anesthesia, on days 0 and 21. Peromyscus are euthanized, using isoflurane anesthesia, on day 30. After infection animals will be monitored for signs of disease daily. Being a reservoir host, Peromyscus do not demonstrate many of the clinical signs commonly seen with Lyme disease infection in animals. Some mild joint edema may be seen, but not to a degree that would affect ambulation.				
5.	Attach or include with the reason(s) for why anesthetics, analgesics and tranquillizers could not be used. (For federally mandated testing, see Item 6 below).			
Anesthetic (isoflurane) is used at the time of infection, for all blood collections and at the time of euthanasia. Analgesics are not used in this study due to concerns that steroids or non-steroidal anti-inflammatory agents may alter the progression of disease and infection. Opioid administration is also associated with immunomodulatory effects, including inhibition of antibody and cellular responses, natural killer cell activity, cytokine expression, chemokine-induced chemotaxis, and phagocytic activity.				
6.	What, if any, federal regulation require this procedure? Cite the agency, the code of Federal Regulations (CFR) title number and the specific section number (e.g. APHIS, 9 CFR 113.102):			
	AgencyCFR			

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Obtained by Rise for Animals.

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1.	Registration	Number: <u>63-R-010</u>	07	-	
2.	Number	199	of animals ι	used in this study.	
3.	Species (cor	nmon name) Hamste	er	of animals used in this study.	
4.	description include, but	of the procedure, bu	ut also explain wha Neurological signs,	ress. Explanations should include a brief at the animal's experience, examples of which may seizures, tremors, paralysis, lethargy, and diarrhea.	
fectomp	tion animals a ounds or vaco e performed o	re monitored for sign cines via the intraven	ns of disease daily. I nous, intraperitonea collected at differen	as a model of human COVID-19 infection. After Hamsters will be treated with test therapeutic al or subcutaneous route. Body weight measurements at time points throughout the studies. Some hamsters test therapeutics.	
corin	ng system has		s project and includ	e, weight loss, lethargy, and difficulty moving. A de the following euthanasia criteria: inappetence, tly closed eyes.	
5.		lude with the reason derally mandated to		netics, analgesics and tranquillizers could not be below).	
nesthetic (ketamine/ xylazine) is used at the time of infection and at the time of euthanasia. Isoflurane is used or all in vivo blood collections. Analgesics are not used in this study due to concerns that if we provide therapy ther than the treatment/compound being tested, we will not be able to evaluate the efficacy of the nerapeutic agent.					
6.		A constraint remove the contract		edure? Cite the agency, the code of Federal ction number (e.g. APHIS, 9 CFR 113.102):	
	Agency_		CFR		

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1.	Registration	Number: 63-R-010	)7			
2.	Number	16	of animals used in this study.			
3.	Species (con	nmon name) <u>Hams</u>	of animals used in this study.			
4.	Explain the procedure producing pain and/or distress. Explanations should include a brief description of the procedure, but also explain what the animal's experience, examples of which may include, but are not limited to: Neurological signs, seizures, tremors, paralysis, lethargy, inappetance, respiratory signs, GI distress, vomiting, and diarrhea.					
subcu	itaneous infec	tion animals will be	ra interrogans as a model of acute human leptospirosis. After monitored for signs of disease. Body weight measurements will be ill occur at the start of the experiment and then every 7 days.			
movii	ng. A scoring s	ystem has been crea	show inappetence, weight loss, lethargy, conjunctivitis and difficulty ated for this project and include the following euthanasia criteria: y/reluctance to move, permanently closed eyes.			
5.			n(s) for why anesthetics, analgesics and tranquillizers could not be esting, see Item 6 below).			
Analg may a immu	esics are not unalled the programme the prog	used in this study du ession of disease and effects, including in	ne of infection, for all blood collections and at the time of euthanasia. e to concerns that steroids or non-steroidal anti-inflammatory agents d infection. Opiod administration is also associated with hibition of antibody and cellular responses, natural killer cell activity, d chemotaxis, and phagocytic acitivity.			
6.			require this procedure? Cite the agency, the code of Federal and the specific section number (e.g. APHIS, 9 CFR 113.102):			
	Agency		CFR			